
B2GD1309

Name Yoshiharu Matsumae

1	1
1.1	1
1.2	SECM, Scanning Electrochemical Microscopy	2
1.2.1	SECM	2
1.2.2	3
1.2.3	4
1.2.4	Generation-Collection	5
1.2.5	5
1.2.6	SECM	6
1.2.7	SECM [3].....	7
1.3	SECM	8
1.3.1	8
1.3.2	11
1.3.3	12
1.3.4	13
1.3.5	14
1.3.6	14
1.4	15
	16
2	21
2.1	21
2.2	21
2.2.1	21
2.2.2	23
2.3	SECM	24
2.4	24
2.4.1	25

4.2.4	52
4.2.5	53
4.2.6	53
4.3	58
4.3.1	58
4.3.2 ES	58
4.3.4	60
4.4	61
	62
5	SECM	65
5.1	65
5.2	66
	68
6	71
6.1	71
6.2	73
	74
7	77
	79

- ' Yoshiharu Matsumae Y. Takahashi, K. Ino, H. Shiku, T. Matsue, Electrochemical monitoring of intracellular enzyme activity of single living mammalian cells by using a double-mediator system, *Anal. Chim. Acta.* , 2014, **842** 20
- ' Yoshiharu Matsumae T. Arai, Y. Takahashi, K. Ino, H. Shiku, T. Matsue, Evaluation of the differentiation status of single embryonic stem cells using scanning electrochemical microscopy *Chem. Commun.* , 2013, **49**, 6498
- ' Yoshiharu Matsumae Y. Takahashi, H. Ida, K. Ino, H. Shiku, T. Matsue, Mediator-free Electrochemical monitoring of Cetuximab-induced Internalization of Epidermal Growth Factor Receptors on Single Cell by Scanning Electrochemical Microscopy, *in preparation*
- ' Toshiharu Arai, T. Nishijo, Y. Matsumae, Y. Zhou, K. Ino, H. Shiku, T. Matsue, Noninvasive Measurement of Alkaline Phosphatase Activity in Embryoid Bodies and Coculture Spheroids with Scanning Electrochemical Microscopy *Anal. Chem.* , 2013, **85**, 9647
- ' Yasufumi Takahashi, K. Ito, X. Wang, Y. Matsumae, H. Komaki, A. Kumatani, K. Ino, H. Shiku, T. Matsue, Nanoscale Cell Surface Topography Imaging using Scanning Ion Conductance Microscopy *Electrochemistry*, 2014, **82** 331
- ' Aiguo Zhang, Y. Matsumae, X. Wang, H. Komaki, A. Kumatani, K. Ino, H. Shiku, T. Matsue, Improving the electrochemical imaging sensitivity of scanning electrochemical microscopy/scanning ion conductance microscopy by using electrochemical Pt deposition, *submitted*

2013 12 p. 751

' 22

' 26

1

1.1

17

19

1953

DNA

19

iPS

iPS

SPM

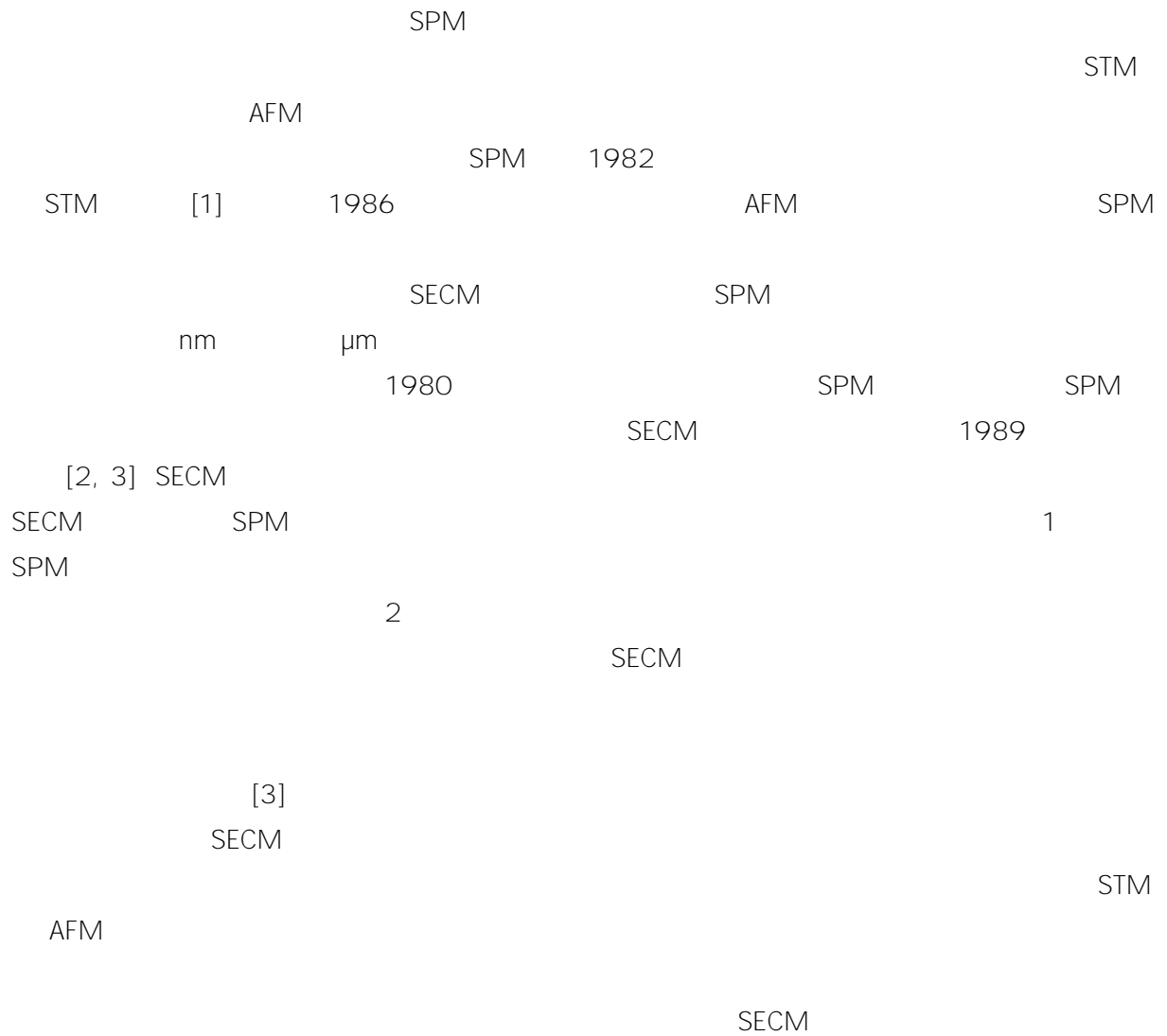
SECM

SECM

1

SECM

1.2 SECM, Scanning Electrochemical Microscopy



1.2.1 SECM

SECM SPM

SECM

Figure

1.1

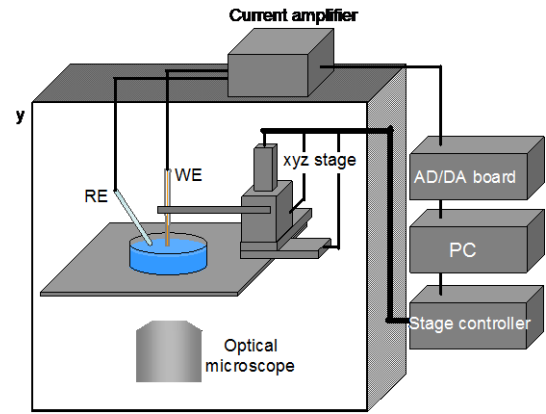


Figure 1.1 SECM system

PC

LabView Visual Basic

1.2.2

SECM

[4,5]

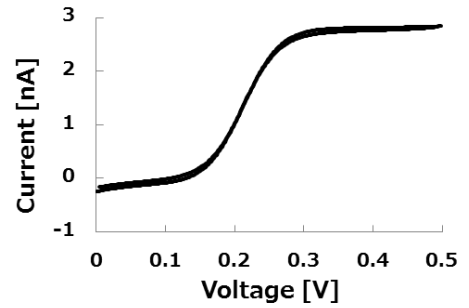


Figure 1.2 Cyclic voltammogram (CV) of microelectrode[8]. Pt microelectrode (radius = 10 m), 1 mM FcCH₂OH in PBS. Scan rate is 20 mV s⁻¹.

i_{tip} , Figure 1.2 1 [4-7]

1

(n: [-], F: =96500 [C/mol], D: [cm²/s], C: [mol/cm³], a: [cm])

Figure 1.3 A
3

1
Figure 1.3 B

S/N

r_g $RG=r_g/a$
4 4.43

RG 2 (1)

RG 10
(1)

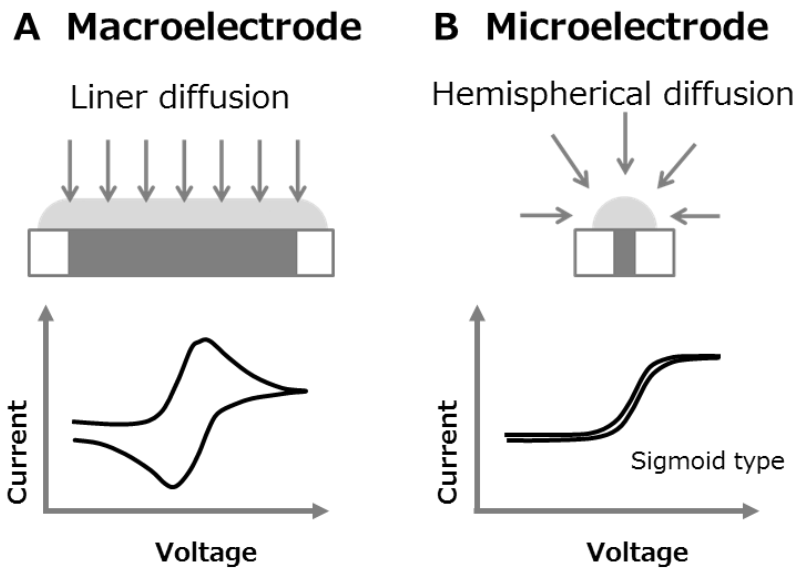


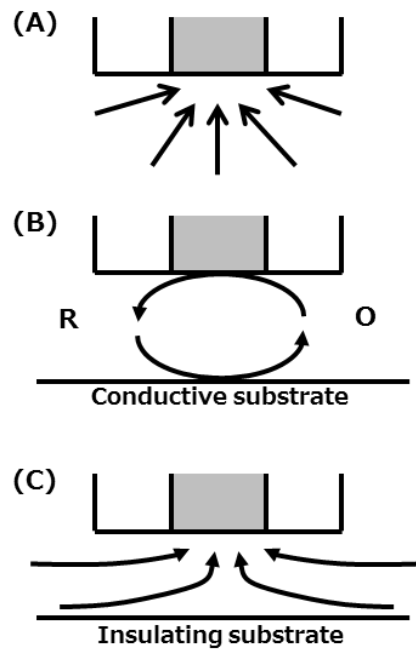
Figure 1.3 Diffusion layer of macroelectrode (A: linear diffusion) and microelectrode (B: hemispherical diffusion) [5].

1.2.3

1

Figure 1.4 A
Figure 1.4 C

Figure 1.4 B



z

1.2.4 Generation-Collection

GC

Figure 1.5

Figure 1.4 Basic principle of SECM[3].

A: hemispherical diffusion

B: positive feedback

C: negative feedback

GC SECM

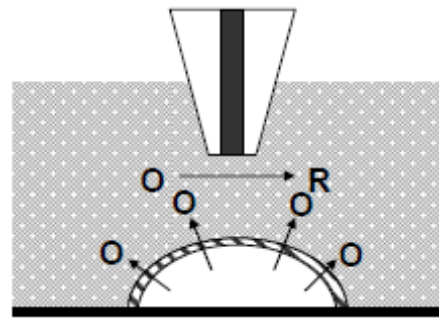


Figure 1.5 GC mode

1.2.5

SECM

constant height

Figure 1.6A

constant distance

Figure 1.6B

XY

SECM

SECM

FB

Z

FB

Z

[9, 10]

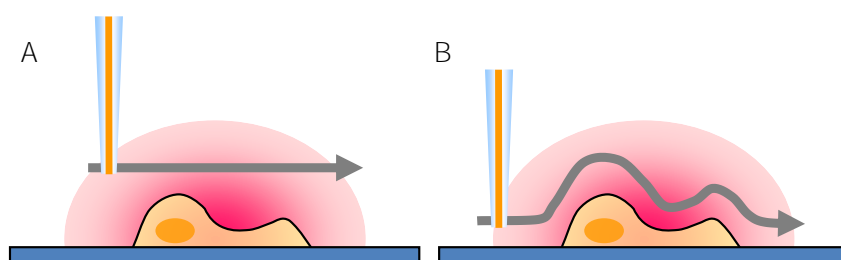


Figure 1.6 (A) constant heightmode, (B) constant distance mode

1.2.6

SECM

[21-30]

[31-33]

[11~15]

[9, 1620]

SECM

150 nm SICM [34,35] SICM Ag/AgCl 50

[33] SECM SECM SICM SECM [32]

AFM

SECM

1.2.7 SECM

[3]

SECM

SECM

3

SECM

2

SECM

7

SECM

SECM
SECM

1.3 SECM

1.3.1

SECM

1

SECM
SECM
GOx

2 3 [3]

Dp

ALP

Figure 1.7

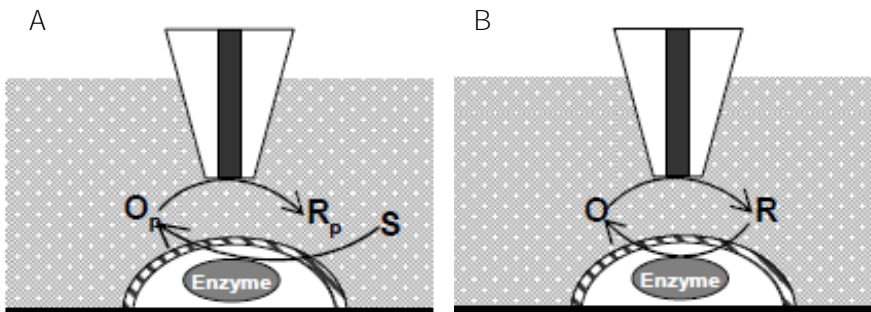
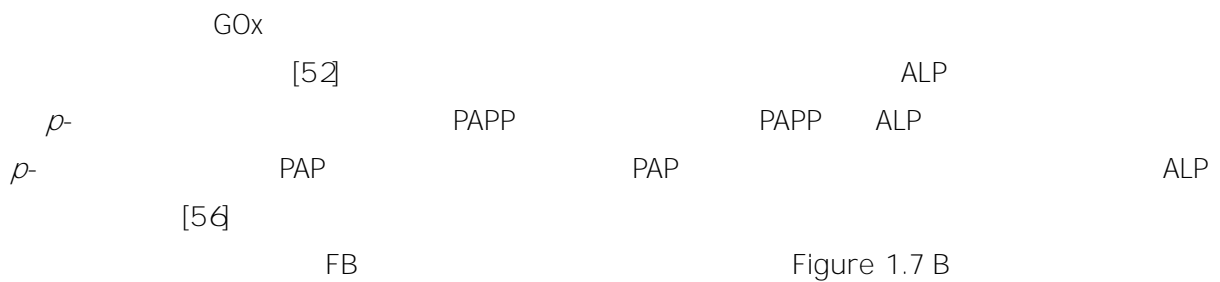


Figure 1.7 Detection principles of enzymes by GC mode (A) or FB mode (B)

GC

Figure 1.7 A



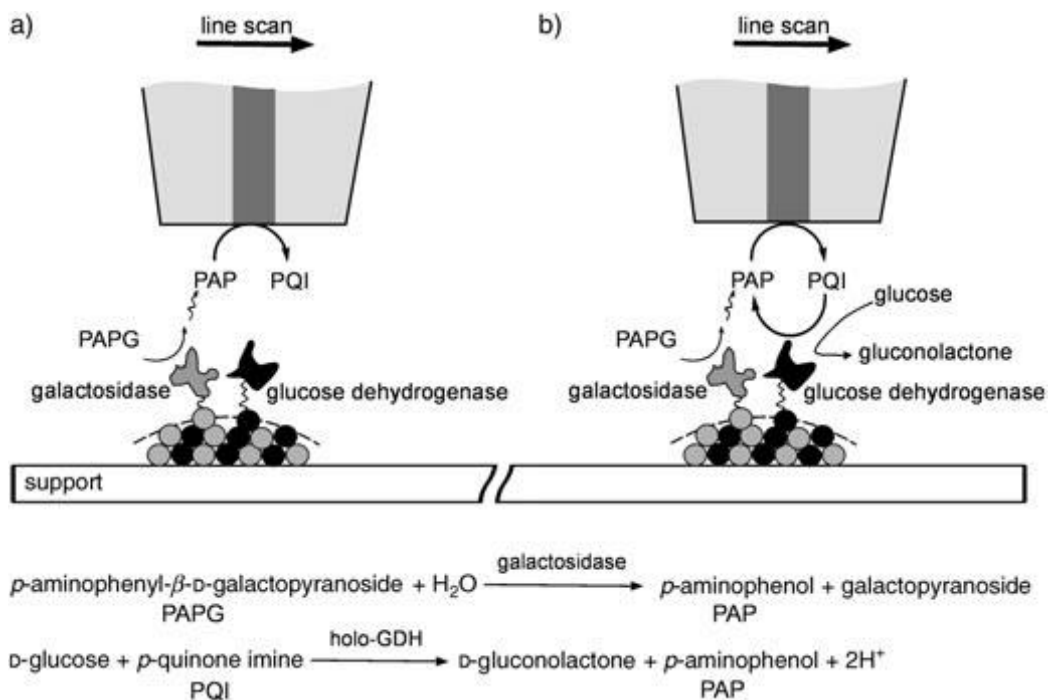


Figure 1.9 Detection by GC mode (A) or both GC mode and FB mode.

Wittstok 2 FB
 GC Figure 1.9 [59, 6Q]
 GDH
 PAPG *p*- D
 Gal PAP PAP
p- PAPI Figure 1.9A PAPI GDH PAP
 PAP Figure 1.9B PAP
 GC 1.8
 [79-82]
 [61]
 ALP [53]
 [83] -GAL
 ALP [84-86]

2 Measurements of enzymatic activities by FB mode.

		FcCOOH	45-47
		(CH ₃) ₂ NCH ₂ Fc	45
		K ₄ [Fe(CN) ₆]	45,48
		hydroquinone	48
		[Os fpy(bpy) ₂ Cl]Cl	46

NADPH	C	NADH	TMPD	48
		NADH	FcCH ₂ OH	49
		H ₂ O ₂	(FcCH ₂ OH) ⁺	50
		NO ₃ ⁻	MV ²⁺	51
Fc: ferrocene fpy: formylpyridine, bpy: bipyridine TMPD: N,N,N',N'-tetramethyl- <i>p</i> -phenylenediamine MV ²⁺ : methylviologen				

3 Measurements of enzymatic activities by GC mode.

	H ₂ O ₂	18,52
	H ⁺	54
	PAPP	56
NAD ⁺	H ⁺	57
NADPH	O ₂ ⁻	58
PAPP: <i>p</i> -aminophenyl phosphate		

1.3.2

1998

SECM

[62 63]

Figure 1.11 SECM

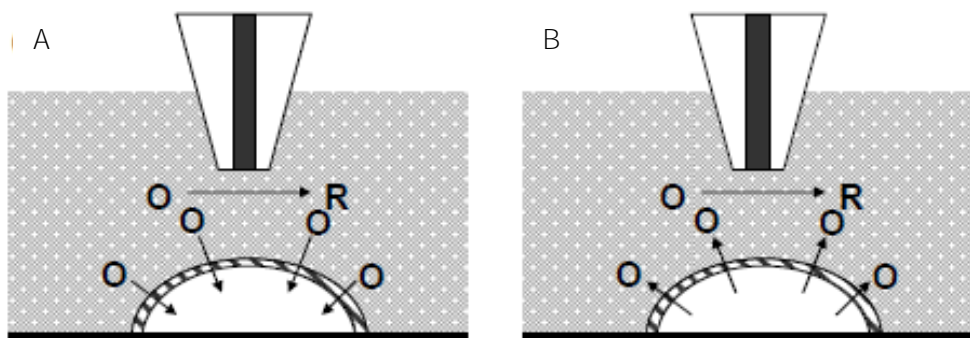
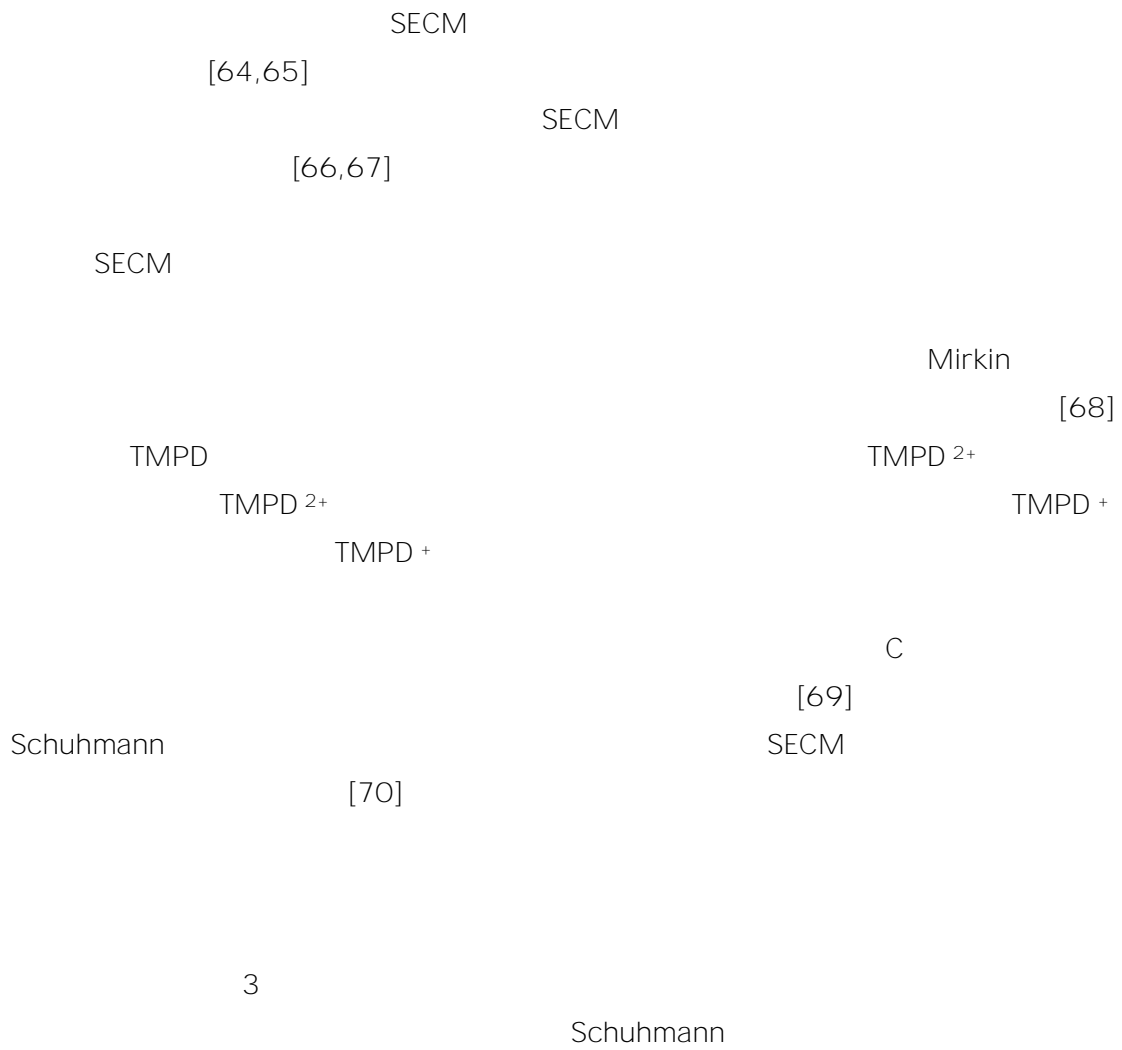


Figure 1.10 A: O₂ consumption by respiration, B: O₂ generation by photosynthesis



1.3.3

SECM

SECM
Ru(NH₃)₆³⁺

[71]

Ru(NH₃)₆³⁺
Figure 1.12 Ru(NH₃)₆³⁺

pH
pH

pH

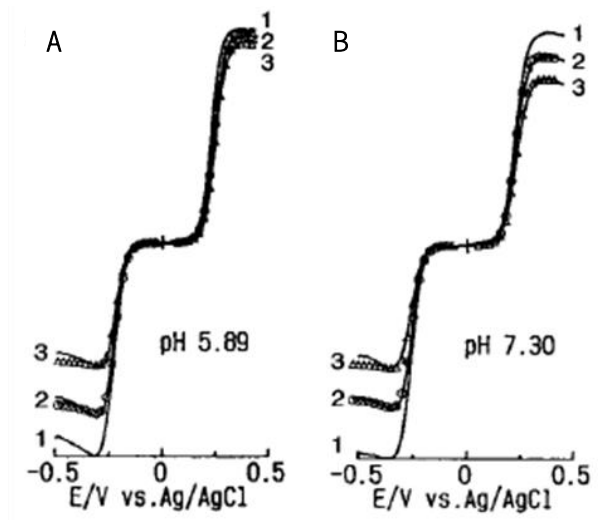


Figure 1.11 Relationship between CV s and membrane-electrode distances. [71]

1: bulk, 2: 51 m, 3: 27 m, A: pH 5.89, B: pH 7.30

Amemiya

[72]

SECM

[73,74]

1.3.4

SECM

[9,22,75-78]

FSCV

[9,22]

FSCV

SECM

[9,22,76,77]

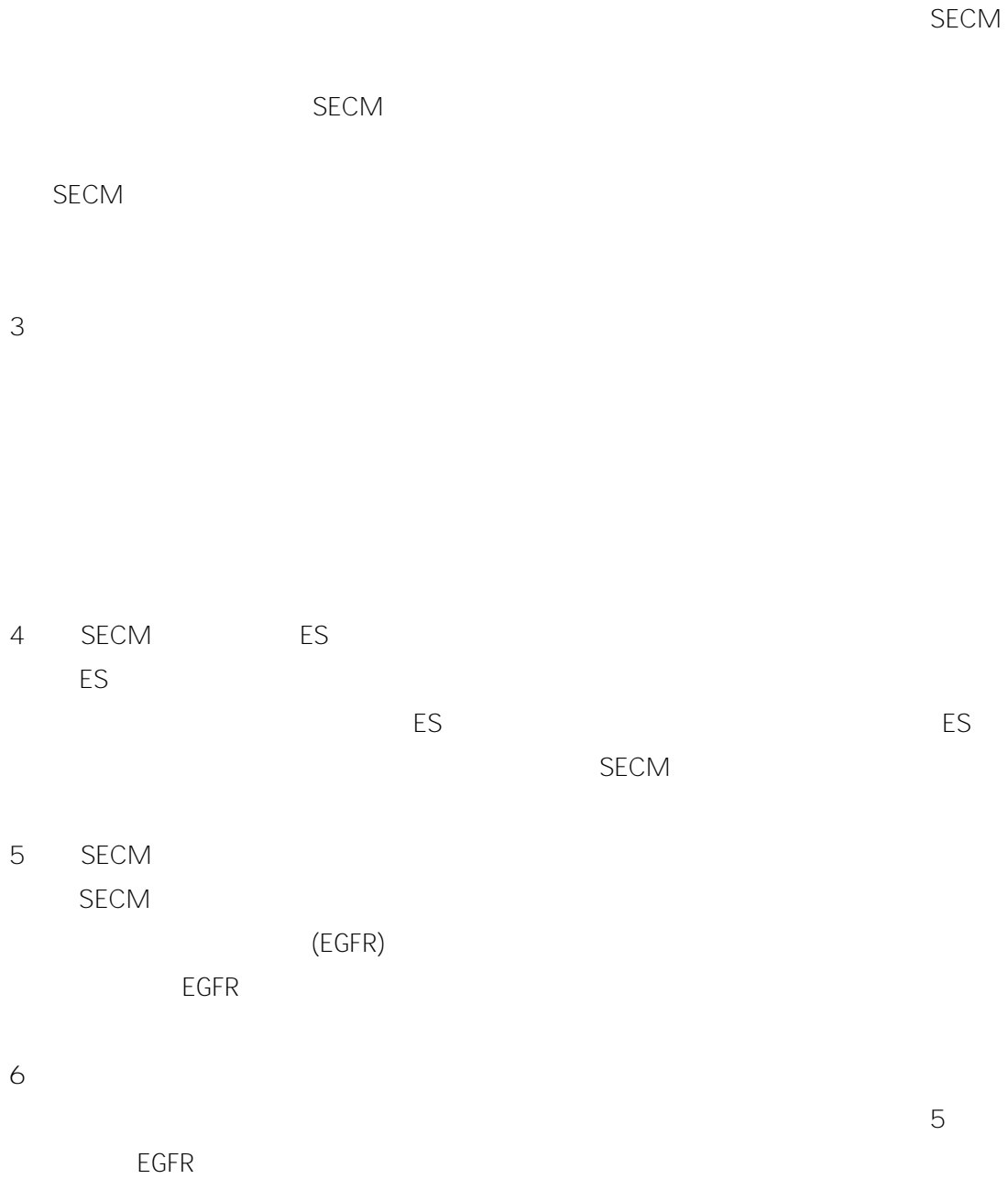
1.3.5

Ru(NH₃)₆³⁺ PC12 [78]
Wipf
SECM
SECM PC12
[9] Takahashi
PC12 [21, 33] Takahashi
[77]

1.3.6

ROS RNS [87, 88]
[89, 91]
[92, 101]
[49, 52, 123] [102, 104, 111, 114-116] [110, 115, 121, 122]
[46, 106-108] [109]
[103, 105] [117-119]
[116, 118, 120]
SECM [124]
[120, 125-130] [113, 131-139]
(100) Ag
[140]

1.4



- 1) G. Binning *et al.*, *Phys. Rev. Lett.* , **49** 57 (1982)
- 2) R. C. Engstorm *et al.*, *Anal. Chem.* , **58** 844 (1986)
- 3) A. J. Bard *et al.*, "Scanning Electrochemical microscopy", Marcel Dekker, Inc., New York (2001)
- 4) 1998
- 5) 2002
- 6) J. L. Amphlett *et al.*, *Anal. Chem.* , **73** 4873 (2001)
- 7) Y. Shao *et al.*, *J. Phys. Chem. B.* , **102** 9915 (1998)
- 8) Y. Shao *et al.*, *Anal. Chem.* , **69** 1627 (1997)
- 9) R. T. Kurulugama *et al.*, *Anal. Chem.* , **77**, 1111 (2005)
- 10) R. Guckenberger *et al.*, *Science*, **266** 1538 (1994)
- 11) A. Kueng *et al.*, *Angew. Chem. Int. Ed.* , **44** 3419 (2005)
- 12) Y. Hirata *et al.*, *Bioelectrochemistry*, **63** 217 (2004)
- 13) D. P. Burt *et al.*, *Nano. Lett.* , **5**, 609 (2005)
- 14) P. S. Dobson *et al.*, *Phys. Chem. Chem. Phys.* , **8**, 3909 (2006)
- 15) R. J. Fasching *et al.*, *Sens. Actuators, B*, **108** 964 (2005)
- 16) D. M. Osbourn *et al.*, *Anal. Chem.* , **77**, 6999 (2005)
- 17) B. R. Horrocks *et al.*, *Anal. Chem.* , **65** 3605 (1993)
- 18) A. Mario *et al.*, *Anal. Chem.* , **73**, 4873 (2001)
- 19) M. Etienne *et al.*, *Electrochem. Commun.*, **6**, 288 (2004)
- 20) C. Gabrielli *et al.*, *Phys. Chem. B.* , **108** 11620(2004)
- 21) Y. Takahashi *et al.*, *Langmuir* , **22** 10229 (2006)
- 22) A. Hengstenberg *et al.*, *Angew. Chem. Int. Ed.* , **40** 905 (2001)
- 23) L. P. Bauermann *et al.*, *Phys. Chem. Chem. Phys.* , **6**, 4003 (2004)
- 24) S. Isik *et al.*, *Angew. Chem. Int. Ed.* , **45** 7451 (2006)
- 25) Y. Lee *et al.*, *Anal. Chem.* , **74**, 3634 (2002)
- 26) D. Oyamatsu *et al.*, *Bioelectrochemistry*, **60** 115 (2003)
- 27) H. Yamada *et al.*, *Anal. Chem.* , **77**, 1785 (2005)
- 28) M. Etienne *et al.*, *Anal. Chem.* , **78** 7317 (2006)
- 29) M. Büchler *et al.*, *Electrochem. Solid-State Lett.*, **3**, 35 (2000)
- 30) M. F. Garay *et al.*, *Phys. Chem. Chem. Phys.* , **6**, 4028 (2004)
- 31) D. J. Comstock *et al.*, *Anal. Chem.* , **82** 1240 (2010)
- 32) Y. Takahashi *et al.*, *J. Am. Chem. Soc.* , **132** 10118 (2010)
- 33) Y. Takahashi *et al.*, *Angew. Chem. Int. Ed.* , **50** 9638 (2011)
- 34) Y. E. Korchev *et al.*, *Biophys. J.* , **73** 653 (1997)
- 35) P. Noval *et al.*, *Nature Methods*, **6**, 279 (2009)

- 36) P. M. Penner *et al.*, *Science*, **250** 1118 (1990)
- 37) M. V. Mirkin *et al.*, *J. Electroanal. Chem.*, **328** 47 (1992)
- 38) J. L. Coners *et al.*, *Anal. Chem.*, **72** 4441 (2000)
- 39) K. Maruyama *et al.*, *Anal. Chem.*, **78** 1904 (2006)
- 40) B. Zhang *et al.*, *Anal. Chem.*, **76** 6229 (2004)
- 41) J. J. Watkins *et al.*, *Anal. Chem.*, **75** 3962 (2003)
- 42) A. Kucernak *et al.*, *J. Phys. Chem. B*, **106** 9396 (2002)
- 43) D. H. Woo *et al.*, *Anal. Chem.*, **75** 6732 (2003)
- 44) N. J. Gray *et al.*, *Analyst*, **125** 889 (2000)
- 45) D. T. Pierce *et al.*, *Anal. Chem.*, **64** 1795 (1992)
- 46) C. Kranz *et al.*, *Electrochim. Acta*, **42** 3105 (1997)
- 47) C. A. Wijayawardhana *et al.*, *Anal. Chem.*, **72** 333(2000)
- 48) D. T. Pierce *et al.*, *Anal. Chem.*, **65** 3598 (1993)
- 49) H. Shiku *et al.*, *Anal. Chem.*, **67**, 312 (1995)
- 50) H. Shiku *et al.*, *Anal. Chem.*, **68** 1276 (1996)
- 51) J. Zaumseil *et al.*, *Anal. Chem.*, **367**, 352 (2000)
- 52) G. Wittstock *et al.*, *Anal. Chem.*, **69**, 5059 (1997)
- 53) Y. Matsumae *et al.*, *Chem. Commun.*, **49**, 6498 (2013)
- 54) B. R. Horrocks *et al.*, *Anal. Chem.*, **65** 1213 (1993)
- 55) B. R. Horrocks *et al.*, *J. Chem. Soc. Faraday Trans.*, **94** 1115 (1998)
- 56) G. Wittstock *et al.*, *Anal. Chem.*, **67**, 3578 (1995)
- 57) Y. N. Antoneko *et al.*, *Arch. Biochem. Biophys.*, **333** 225 (1996)
- 58) C. E. M. Berger *et al.*, *J. Endocrin.*, **158** 311 (1998)
- 59) C. Zhao *et al.*, *Angew. Chem. Int. Ed.*, **43** 4170 (2004)
- 60) F. M. Boldt *et al.*, *Anal. Chem.*, **76** 3473 (2004)
- 61) K. Nagamine *et al.*, *Electroanal.*, **23** 1168 (2011)
- 62) T. Yasukawa *et al.*, *Chem. Lett.*, **8**, 767 (1998)
- 63) T. Yasukawa *et al.*, *Anal. Chem.*, **71**, 4637 (1999)
- 64) T. Kaya *et al.*, *Biosens. Bioelectron.*, **18**, 1379 (2003)
- 65) M. Nishizawa *et al.*, *Langmuir*, **18** 3645 (2002)
- 66) H. Shiku *et al.*, *Anal. Chem.*, **73** 3751 (2001)
- 67) T. Saito *et al.*, *Analyst*, **131**, 1006 (2006)
- 68) W. Feng *et al.*, *Anal. Chem.*, **75** 4148 (2003)
- 69) A. J. Bard *et al.*, *Biosens. Bioelectron.*, **22** 461 (2006)
- 70) M. Nebel *et al.*, *Angew. Chem. Int. Ed.*, **52** 1 (2013)
- 71) H. Yamada *et al.*, *Biochem. Biophys. Res. Commun.*, **180** 1330 (1991)
- 72) T. Yasukawa *et al.*, *Biochim. Biophys. Acta*, **1369** 152 (1998)

- 73) J. Guo *et al.*, *Anal. Chem.* , **77**, 2147 (2005)
- 74) J. Kim *et al.*, *J. Am. Chem. Soc.* , **135** 2321 (2013)
- 75) T. K. Chen *et al.*, *Anal. Chem.* , **66** 3031 (1994)
- 76) Y. Takahashi *et al.*, *Angew. Chem. Int. Ed.* , **50** 9638 (2011)
- 77) Y. Takahashi *et al.*, *Proc. Natl. Acad. Sci. U. S. A.* , **109** 1154 (2012)
- 78) J. M. Liebetrau *et al.*, *Anal. Chem.* , **75** 563 (2003)
- 79) Y. Takahashi *et al.*, *Anal. Chem.* , **81**, 2785 (2009)
- 80) Y. Takahashi *et al.*, *Phys. Chem. Chem. Phys.* , **13**, 16569 (2011)
- 81) S. Kasai *et al.*, *Anal. Chem.* , **72** 5761 (2000)
- 82) H. Shiku *et al.*, *J. Electroanal. Chem.* , **438** 1 (1997)
- 83) Y. Xue *et al.*, *Anal. Chem.* , **82** 7112 (2010)
- 84) T. Kaya *et al.*, *Chem. Commun.* , 248 (2004)
- 85) N. Matsui *et al.*, *Biosens. Bioelectron.* , **21**, 1202 (2006)
- 86) Y. S. Torisawa *et al.*, *Anal. Chem.* , **78** 7625 (2006)
- 87) J. Mauzeroll *et al.*, *Proc. Natl. Acad. Sci. U. S. A.* , **101**, 7862 (2004)
- 88) J. Mauzeroll *et al.*, *Proc. Natl. Acad. Sci. U. S. A.* , **101**, 17582 (2004)
- 89) C. Amatore *et al.*, *ChemBioChem* , **7**, 653 (2006)
- 90) C. Amatore *et al.*, *ChemBioChem* , **9**, 1472 (2008)
- 91) C. Amatore *et al.*, *Anal. Chem.* , **82** 1411 (2010)
- 92) E. R. Scott *et al.*, *Anal. Chem.* , **65** 1537 (1993)
- 93) E. R. Scott *et al.*, *J. Membrane* , **58** 71 (1991)
- 94) S. Nugues *et al.*, *J. Electroanal. Chem.* , **408** 125 (1996)
- 95) B. D. Bath *et al.*, *Anal. Chem.* , **70** 1047 (1998)
- 96) J. V. Macpherson *et al.*, *J. Chem. Soc. Faraday Trans.* , **91**, 1407 (1995)
- 97) J. V. Macpherson *et al.*, *Langmuir* , **11**, 3959 (1995)
- 98) J. V. Macpherson *et al.*, *Biophys. J.* , **73** 2771 (1997)
- 99) E. R. Scott *et al.*, *Solid State Ion.* , **53** 176 (1992)
- 100) E. R. Scott *et al.*, *Pharm. Res.* , **10**, 1699 (1993)
- 101) E. R. Scott *et al.*, *J. Invest. Dermatol.* , **104** 142 (1995)
- 102) D. H. Craston *et al.*, *J. Electrochem. Soc.*, **135** 785 (1988)
- 103) O. E. Hüsser *et al.*, *J. Vac. Sci. Technol. B* , **6**, 1873 (1988)
- 104) O. E. Hüsser *et al.*, *J. Electrochem. Soc.*, **136** 3222 (1989)
- 105) F. Forouzan *et al.*, *J. Phys. Chem. B* , **101**, 10876 (1997)
- 106) Y. M. Wu *et al.*, *J. Electrochem. Soc.*, **136** 885 (1989)
- 107) C. Kranz *et al.*, *Adv. Mater.* , **7**, 38 (1995)
- 108) C. Kranz *et al.*, *Adv. Mater.* , **7**, 568 (1995)
- 109) H. Sugimura *et al.*, *Electroanal. Chem.*, **361** 57 (1993)

- 110) G. Wittstock *et al.*, *Electroanal.*, **9**, 746 (1997)
- 111) D. Mandler *et al.*, *J. Electrochem. Soc.*, **137**, 1079 (1990)
- 112) D. Mandler *et al.*, *J. Electrochem. Chem.*, **136** 3134 (1989)
- 113) J. W. Still *et al.*, *J. Electrochem. Soc.*, **144** 2657 (1997)
- 114) S. Meltzer *et al.*, *J. Electrochem. Soc.*, **142** L82 (1995)
- 115) K. Borgwarth *et al.*, *Ber. Bunsenges. Phys. Chem.*, **99** 1421 (1995)
- 116) I. Shohat *et al.*, *J. Electrochem. Soc.*, **141**, 995 (1994)
- 117) D. Mandler *et al.*, *J. Electrochem. Soc.*, **137**, 2468 (1990)
- 118) I. Turyan *et al.*, *Adv. Mater.*, **12** 330 (2000)
- 119) S. Meltzer *et al.*, *J. Chem. Soc. Faraday Trans.*, **91**, 1019 (1995)
- 120) J. V. Macpherson *et al.*, *J. Am. Chem. Soc.*, **118**, 6445 (1996)
- 121) K. Borgwarth *et al.*, *Adv. Mater.*, **11**, 1221 (1999)
- 122) J. Zhou *et al.*, *J. Electrochem. Soc.*, **144** 1202 (1997)
- 123) H. Shiku *et al.*, *Langmuir*, **13**, 7239 (1997)
- 124) P. R. Unwin *et al.*, *J. Phys. Chem.*, **96** 5035 (1992)
- 125) J. V. Macpherson *et al.*, *J. Chem. Soc. Faraday Trans.*, **89**, 1883 (1993)
- 126) J. V. Macpherson *et al.*, *J. Phys. Chem.*, **98** 1704 (1994)
- 127) J. V. Macpherson *et al.*, *J. Phys. Chem.*, **98** 11764 (1994)
- 128) J. V. Macpherson *et al.*, *J. Phys. Chem.*, **99**, 3338 (1995)
- 129) J. V. Macpherson *et al.*, *J. Phys. Chem.*, **99**, 14824 (1995)
- 130) J. V. Macpherson *et al.*, *J. Phys. Chem.*, **100** 19475 (1996)
- 131) N. Casillas *et al.*, *J. Electrochem. Soc.*, **140** L142 (1993)
- 132) N. Casillas *et al.*, *J. Electrochem. Soc.*, **141**, 636 (1994)
- 133) L. F. Garfias-Mesias *et al.*, *J. Electrochem. Soc.*, **145** 2005 (1998)
- 134) Y. Zhu *et al.*, *J. Electrochem. Soc.*, **144** L43 (1997)
- 135) D. E. Williams *et al.*, *J. Electrochem. Soc.*, **145** 2664 (1998)
- 136) D. O. Wipf *et al.*, *Colloids Surf. A: Physicochem. Eng. Asp.*, **93** 251 (1994)
- 137) N. Casillas *et al.*, *J. Electrochem. Soc.*, **142** L16 (1995)
- 138) P. James *et al.*, *J. Electrochem. Soc.*, **143** 3853 (1996)
- 139) J. L. Luo *et al.*, *J. Electroanal. Chem.*, **326** 51 (1992)
- 140) P. R. Unwin *et al.*, Abstracts of the 193rd Meeting of The Electrochemical Society, n**980**(1998)

Pt
M NaOH

0.01

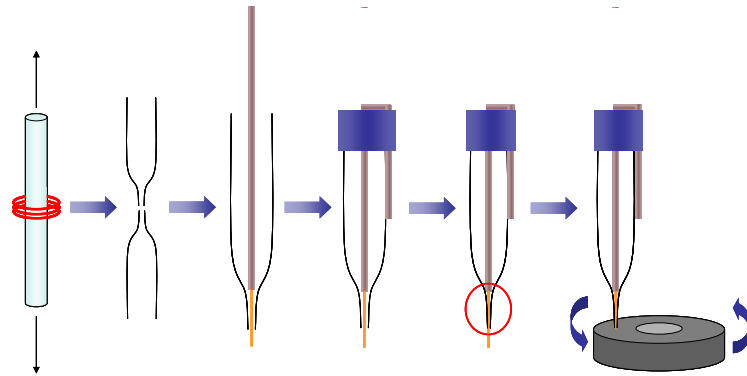


Figure 2.1 Fabrication procedure of Pt disk-type microelectrode.

j_{bulk}

1

j_{bulk} [A] n F [C/mol] D
[cm²/sec] C [mol/cm³] a [cm]
FcCH₂OH K₄Fe(CN)₆

Figure 2.2

Pt RG 2 4.43
10 am
Fe(CN)₆⁴⁻ 6.5³ 10⁻⁶ [cm² s⁻¹]

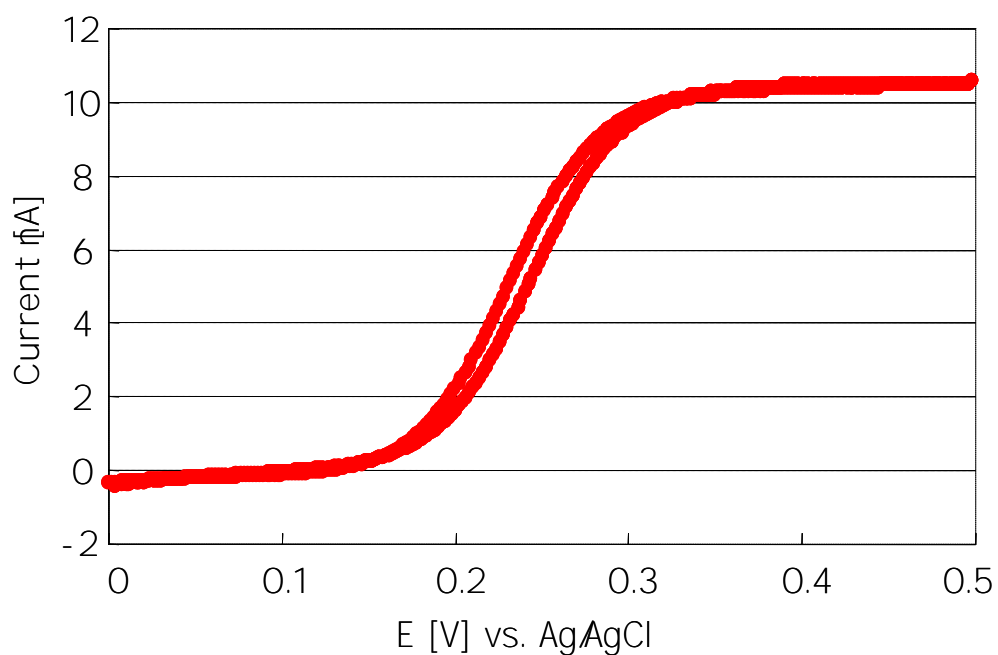
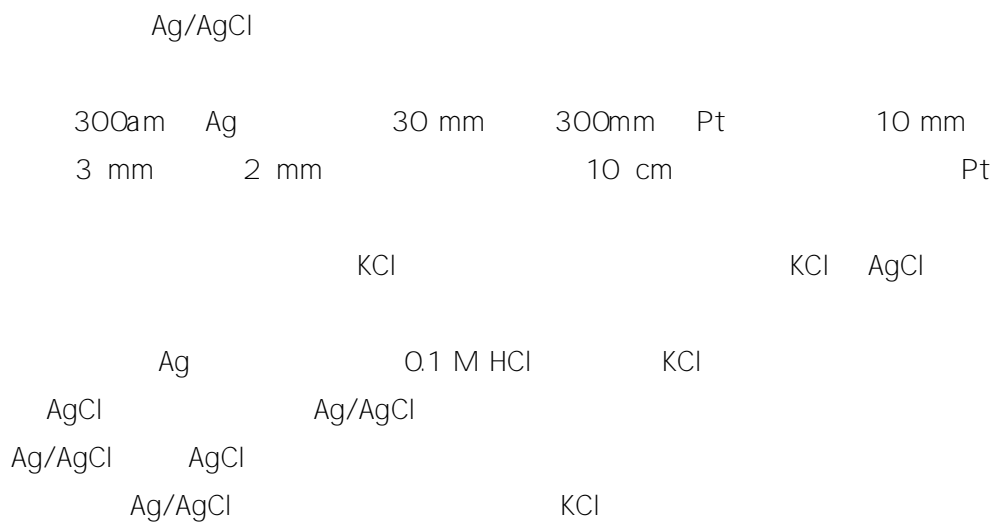


Figure 2.2 Cyclic voltammograms by a fabricated Pt disk-type microelectrode (radius = 10 μm). Measurement solution contains 4 mM $\text{Fe}(\text{CN})_6^{4-}$. The scan rate was 20 mV vs. Ag/AgCl.

2.2.2



2.3

SECM

Figure 2.3
2
RE
DA
AD
Faraday cage

SECM
WE
Ag/AgCl
Current amplifier
PC
HOKUTO DENKO

Pt
PC
427 current amplifier
KEITHLEY
SECM
xyz

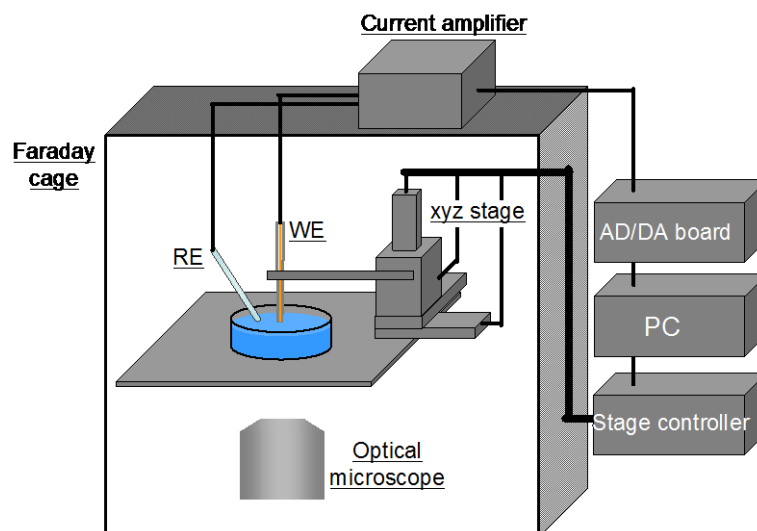


Figure 2.3 Schematic diagram of SECM system

2.4

MCV-B131F

2.4.1

4

HeLa

ES

: Strain 129/ SVEV DS Pharma Biomedical Co. , Ltd

A431

MCF -7

2.4.2

RPMI -1640 Gibco
500mL RPMI -1640 50mL FBS Gibco 5 mL
/
5000 U/mL⁻¹ 5000 µg/mL⁻¹
37°C 56°C 30 min RPMI -1640
4°C
0.25 -EDTA Gibco
1.0 mL
PBS -
8.00 gL⁻¹ NaCl 0.20 gL⁻¹ KCl 1.54 gL⁻¹ NaHPO₄ 12H₂O 0.02 gL⁻¹
KH₂PO₄ KT -2346 121°C 20 min
- Ca²⁺ Mg²⁺
5 10 50mL Falcon
25 250mL Falcon
3.0mL Falcon
15 50 mL IWAKI
C-Chip DHC -BO2 Digital Bio
70%
70%
TE300 Nikon

85°C

2.4.3

ES

HeLa	A431	MCF -7	APM -30D	RPMI -1640
------	------	--------	----------	------------

37°C 5% CO₂

Utrapure water with 0.1 % Gelatin (Millipore, Cat. ES-006-B) 25 cm² 2 mL

4

2 mL PBS(-) PBS

4°C

ES 1000 U mL⁻¹ of mouse leukemia inhibitory factor (mLIF) 1 mM

-mercaptoethanol Stem Medium DS Pharma Biomedical. Co., Ltd., Tokyo, Japan

1 µM ATRA mLIF ES

[1, 2]

2.4.4

HeLa A431 MCF -7

80%

37

PBS - 2

0.25% -EDTA 1 mL 37°C 5%CO₂

CT15D

1500

rpm 5 min

C-Chip

37°C 5%CO₂

ES

37°C

4°C

PBS(-) 5 mL

PBS(-)

Accutase Solution (Millipore, Cat. R-SCR005)

5 mL

5min

10

. (

)

5 mL

15 mL

10

4 °C 100g 3 min

5 mL

C-Chip

1.0³ 10⁶ cells / 5 ml

25 cm²

37

5% CO₂ 95% Air

ES

2.4.5

1.0³ 10⁶ cells mL⁻¹

-80°C

2.4.6

2

37°C

37

1500 rpm 5 min

37°C 5%CO₂

2.5

polydimethylsiloxane PDMS

PDMS

PDMS

Figure 2.5

S1225 MATSUNAMI 2 3 3 4 4 5 5 6
6 6-Nonafluorohexyltrichorosilane 20al
PDMS (184W/C)
1000 rpm 30 80 °C 1
100am PDMS
PDMS CO₂
PDMS 1

PDMS 70%
 PDMS
 1.0 × 10⁶ cells mL⁻¹ 100 μL
 37°C 5% CO₂ 2

80-90%
 37°C 5% CO₂ 1
 PDMS
 PDMS

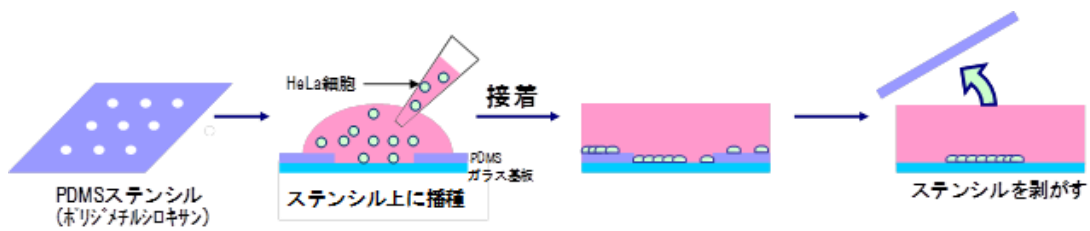


Figure 2.4 Manufacture method of patterned cells by PDMS stencil.

300 μm 100 μm
 Figure 2.6 HeLa

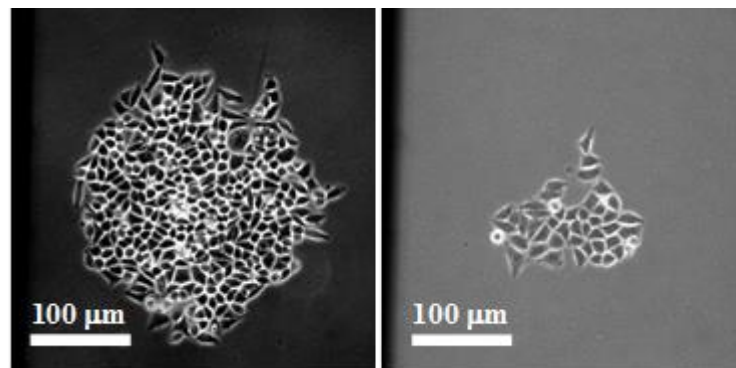


Figure 2.5 Patterned HeLa cells by PDMS stencil.

[1] M. Kim, A. Habiba, J.M. Doherty, J.C. Mills, R.W. Mercer, J.E. Huettner, Regulation of mouse embryonic stem cell neural differentiation by retinoic acid, *Developmental biology*, 328

(2009) 45471.

[2] R. Martin-Ibanez, N. Urban, S. Sergent -Tanguy, J.R. Pineda, N. Garrido -Clua, J. Alberch, J.M. Canals, Interplay of leukemia inhibitory factor and retinoic acid on neural differentiation of mouse embryonic stem cells, *J. Neurosci.*, 85 (2007) 2682701.

[3] 2008

[4] 2008

[5] 2009

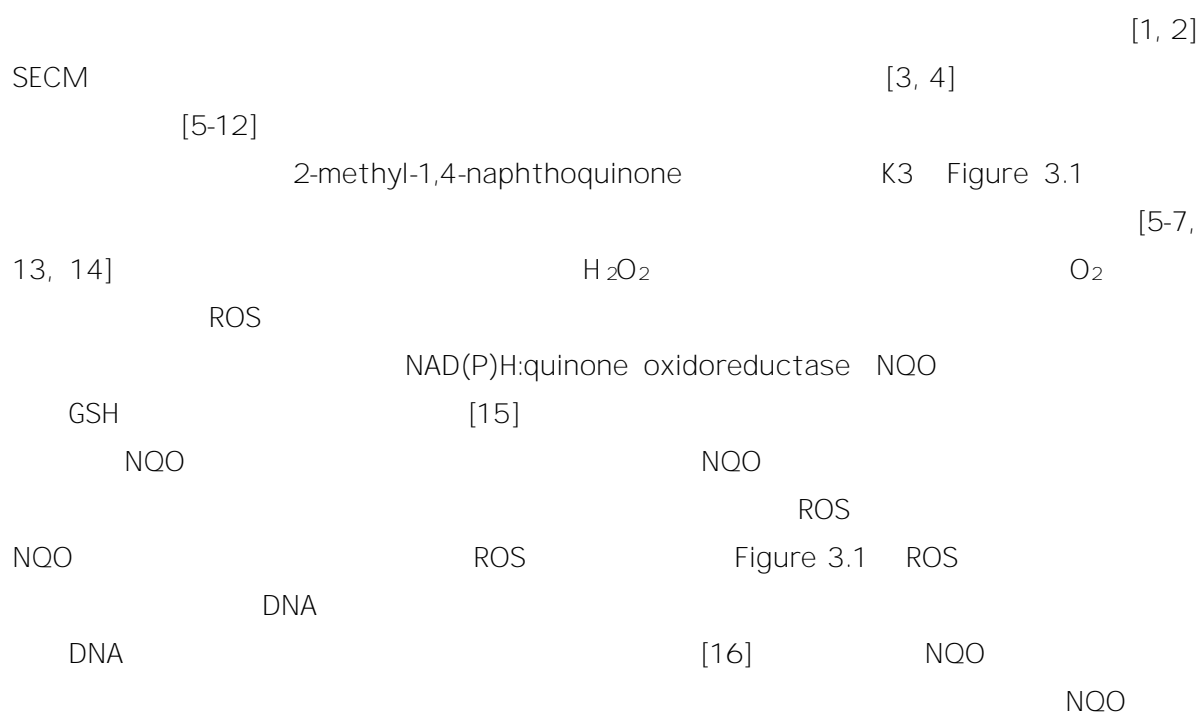
[5] 20013

[4] 20013

3

$K_3Fe(CN)_6$ 2
SECM

3.1



1
GSH
Bard
GSH
GSH
K₃Fe(CN)₆
NQQ
Figure 3.2 [13, 14, 17] Pt
K₃Fe(CN)₆
Fe(CN)₆⁴⁻

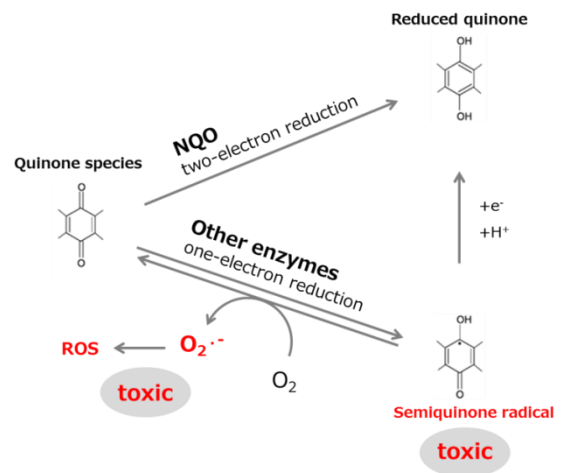


Figure 3.1 Schematic illustration of the reduction of quinone species within the cells. NAD(P)H:quinone oxidoreductase (NQO) can reduce quinone species by two-electron reduction without the generation of reactive oxygen species (ROS) and semiquinone radicals which lead to cytotoxicity.

Nagamine [7]
SECM
K₃Fe(CN)₆
ROS
GSH
SECM
NQO
SECM
NQO

3.2

3.2.1

D-2-methyl-1,4-naphthoquinon KCl NaCl Na₂HPO₄ · 12H₂O MgCl₂ · 6H₂O K₃Fe(CN)₆

N-2-hydroxyethylpiperazine-N'-ethanesulfonic acid HEPES
 Cell Counting Kit -8 OxiSelect™ ROS Assay Kit CELL BIOLABS, INC. GSH -Glo™
 Glutathione Assay Promega HEPES buffer 150 mM NaCl 4.2 mM KCl 2.7 mM MgCl₂
 1.0 mM Na₂HPO₄ 11.2 mM D-glucose 10 mM HEPES

3.2.2 SECM

NQO

Figure 3.2

[2, 5, 18]

NQO

Fe(CN)₆³⁻

Fe(CN)₆⁴⁻

Fe(CN)₆^{3-/4-}

Fe(CN)₆⁴⁻

NQO

NQO

SECM

20 μm Pt

0.5 V vs.

Ag/AgCl

0.5 V vs. Ag/AgCl

Fe(CN)₆⁴⁻

1-line

scan

1-line scan

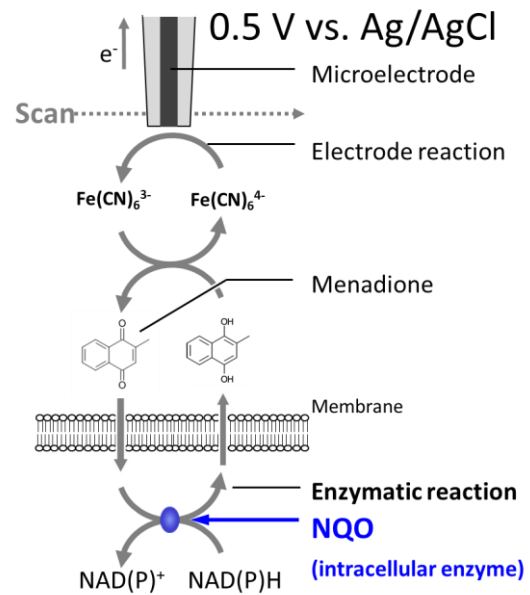


Figure 3.2 Principle of electrochemical detection of NAD(P)H:quinone oxidoreductase(NQO) activity in a HeLa cell. Fe(CN)₆³⁻ and menadione were added in the measurement solution.

3.2.3 NQO

NQO

SECM

[19]

NQO

Fick

flux

